FORM PTO-1390 U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY DOCKET NUMBER **DAVI-0005**
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371		U.S. APPLICATION NO. (if known see 37 C F.R. 1 5) 09/787082
INTERNATIONAL APPLICATION NO. **PCT/AU99/00769**	INTERNATIONAL FILING DATE **14 September 1999 (14.09.99)**	PRIORITY DATE CLAIMED **14 September 1998 (14.09.98)**

TITLE OF INVENTION **CYCLISED CONOTOXIN PEPTIDES**

APPLICANT(S) FOR DO/EO/US **David James CRAIK, Norelle Lee DALY, and Katherine Justine NIELSEN**

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. X This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. _ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. X This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. X A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.

5. X A copy of the International Application as filed (35 U.S.C. 371(c)(2)).
   a. _ is transmitted herewith (required only if not transmitted by the International Bureau).
   b. X has been transmitted by the International Bureau.
   c. _ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. _ A translation of the International Application into English (35 U.S.C. 371(c)(2)).

7. X Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
   a. _ are transmitted herewith (required only if not transmitted by the International Bureau).
   b. _ have been transmitted by the International Bureau.
   c. _ have not been made; however, the time limit for making such amendments has NOT expired.
   d. X have not been made and will not be made.

8. _ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).

9. _ An oath or declaration of the inventor(s) 35 U.S.C. 371(c)(4).

10. _ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern other document(s) or information included:

11. _ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.

12. _ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.

13. _ A FIRST preliminary amendment.
    _ A SECOND or SUBSEQUENT preliminary amendment.

14. _ A substitute specification.

15. _ A change of power of attorney and/or address letter.

16. X Other items or information:
    - **A copy of the Published PCT Application by WIPO under No. WO 00/15654, including the Search Report.**
    - **A copy of the International Preliminary Examination Report. including sheets the replacement sheets 30 and 32 of the claims which were submitted under Article 34 on June 8, 2000.**
    - **Sequence listing in computer readable and written form with required statement.**

EXPRESS MAIL Mailing Label No. **EL567 667 894US**
Date of Deposit: **March 14, 2001**
I hereby certify that this paper or fee is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231
MAILER **Bob Inforzato**
SIGNATURE B. Inforzato

U.S. APPLICATION NO. (if known 37 C.F.R. 1 5)	INTERNATIONAL APPLICATION NO.	ATTORNEY DOCKET NUMBER
09/787082	**PCT/AU99/00769**	**DAVI-0005**

17. x The following fees are submitted:

	CALCULATIONS	PTO USE ONLY
Basic National Fee (37 CFR 1.492(a)(1) - (5)):		
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO.....**$1,000.00**		
International preliminary examination fee (37 CFR 1.482 not paid to USPTO but International Search Report has been prepared by the EPO or JPO.....**$860.00**		
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO.....**$710.00**		
International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4).....**$690.00**		
International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4).....**$100.00**		
ENTER APPROPRIATE BASIC FEE AMOUNT =	**$1000.00**	
Surcharge of $130.00 for furnishing the oath or declaration later that _ 20 _ 30 months from the earliest claimed priority date (37 CFR 1.492(e)).	$	

Claims	Number Filed	Number Extra	Rate		
Total claims	**18** - 20 =	**0**	X $18.00	$ 0	
Independent Claims	**13**- 3 =	**10**	x $80.00	$800.00	
Multiple dependent claims(s) (if applicable)			+ $270.00	$	
			TOTAL OF ABOVE CALCULATIONS =	**$1800.00**	
___ Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by ½.				$	
			SUBTOTAL =	**$1800.00**	
Processing fee of **$130.00** for furnishing the English translation later the _ 20 _30 months from the earliest claimed priority date (37 CFR 1.492(f)). +				$	
			TOTAL NATIONAL FEE =	**$1800.00**	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). **$40.00** per property +					
			TOTAL FEES ENCLOSED =	**$1800.00**	
			Amount to be: refunded	$	
			charged	$	

a. X 2 checks totalling $1800 One in the amount of **$ 1660.00 and the other in the amount of $140.00** to cover the above fee is enclosed.

b. _ Please charge my Deposit Account No. 23-3050 in the amount of $_____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. X The Commissioner if hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 23-3050. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:
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33,229
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CYCLISED CONOTOXIN PEPTIDES

This invention relates to novel peptides and derivatives thereof, in particular to a range of cyclic peptides useful in the therapeutic treatment of humans. The invention also relates to pharmaceutical compositions comprising these peptides, methods for making the peptides and the use of these peptides in the prophylaxis or treatment of conditions or diseases in humans.

The marine nails of the genus *Conus* (cone snails) use a sophisticated biochemical strategy to capture their prey. As predators of either fish, worms or other molluscs, the cone snails inject their prey with venom containing a cocktail of small bioactive peptides. These toxin molecules, which are referred to as conotoxins, interfere with neurotransmission by targeting a variety of receptors and ion-channels. They typically contain 12-30 amino acids arranged in linear sequence. The venom from any single *Conus* species may contain more than 100 different peptides. The conotoxins are divided into classes on the basis of their physiological targets. To date, ten classes have been described. The ω-conotoxin class of peptides target and block voltage-sensitive Ca^{2+}-channels inhibiting neurotransmitter release. The α-conotoxins and ψ-conotoxins target and block nicotinic ACh receptors, causing ganglionic and neuromuscular blockade. Peptides of the μ-conotoxin class act on voltage-sensitive Na^{+}-channels and block muscle and nerve action potentials. The δ-conotoxins target and delay the inactivation of voltage-sensitive Na^{+}-channels enhancing neuronal excitability. The κ-conotoxin class of peptides target and block voltage-sensitive K^{+}-channels, and these may also cause enhanced neuronal excitability. The conopressins are vasopressin receptor antagonists and the conantokins are NMDA receptor antagonists. Recently, the prototype of a new γ-conotoxin class was described, which targets a voltage-sensitive nonspecific cation channel, and of a new σ-conotoxin class, which antagonises the $5HT_3$ receptor.

Most conotoxin peptides contain either four (4) or six (6) cysteine residues which are bonded in pairs to form either two (2) or three (3) disulfide bonds respectively. As indicated above they bind to a range of different ion-channels in mammals, and accordingly they have several potential therapeutic applications, including pain relief and neuroprotection in humans. However, in general peptides have several difficulties associated with their use as drugs,

including generally poor bioavailability, susceptibility to cleavage by proteases, and unwanted side effects.

One conotoxin, MVIIA, is currently in clinical trial for the treatment of intractable pain and for neuroprotection following stroke. In the former indication the route of administration is restricted to intrathecal infusion into the spinal cord because of some of the abovementioned difficulties.

The present invention is based on the finding that cyclisation of the peptide backbone of conotoxins to produce non-natural analogues results in new molecules which can retain the therapeutic activity of the non-cyclised peptide.

Accordingly in a first aspect the present invention provides a cyclised conotoxin peptide.

These cyclised conotoxins have improved properties relative to their "linear" conotoxin counterparts. The improved properties may include the following:

1. Resistance to cleavage by proteases.
2. High chemical stability.
3. An additional "handle" on the molecule which does not interfere with the primary biological effect of the conotoxin, but provides a place for functionalising the molecule to improve biophysical properties or, in some cases, reduce side effects.
4. Improved bioavailability.

The conotoxin peptide may be any conotoxin peptide which is capable of being cyclised. It may be a naturally occurring conotoxin peptide, or a derivative thereof. Preferably the conotoxin peptide is one which, in its non-cyclised form, has an activity associated with the therapeutic treatment of mammals, such as humans. Since the cyclisation of the peptide has the potential to alter the activity of the peptide, or introduce new activities, it is possible that some cyclised conotoxin peptides may have improved therapeutic properties relative to "linear" conotoxins.

Examples of suitable linear naturally occurring conotoxins and derivatives thereof which may be cyclised according to the present invention include those described in Olivera, B.M. *et al.*, 1991; Myers, R.A. *et al.*, 1993; Hopkins, C. *et al.*, 1995; Olivera, B.M. *et al.*, 1990. Preferably the conotoxins are selected from the ω-class, which have characteristic three disulphide bonds forming a "cystine knot", although other classes of conotoxins may also be cyclised.

Examples of suitable naturally occurring ω-conotoxin peptides include MVIIA, GVIA, SVIB, SVIA, TVIA, MVIIC, GVIIA and GVIIB.

The conotoxin peptides have a characteristic folding pattern which is based on the number of disulphide bonds, and the location on the peptide of the cysteine residues which participate in the disulphide bonding pattern. Where there are three disulphide bonds there is the potential for the peptide to form a cystine knot. A cystine knot occurs when a disulphide bond passes through a closed cyclic loop formed by two disulphide bonds and amino acids in the peptide chain. The cyclisation of a conotoxin having a cystine knot produces a particularly stable peptide structure. As well as being present in the class of omega-conotoxins, Nielson, *et al.*, 1996, the cystine knot exists in other classes including, K^+ channel blockers (eg conotoxin PVIIA; Scanlon *et al.*, 1997) and Na channel blockers (eg conotoxin GS; Hill et al., 1997).

Preferred conotoxin peptides are those in which, in their folded form, have N- and C-termini which are located in close proximity. The proximity of termini is illustrated above for MVIIA and PVIIA. In conotoxin GS the N and C termini are further apart, but the C terminus contains a flexible tail which can readily alter conformation to approach the N terminus.

The cyclic conotoxin peptides according to the present invention will generally consist of a conotoxin peptide in which the N- and C-termini are linked via a linking moiety, although in some cases it may be possible to directly connect the N- and C-termini of a naturally occurring conotoxin peptide or derivative thereof without the need for a linking moiety. The linking moiety, if present, may be a peptide linker such that cyclisation produces an amide-

cyclised peptide backbone. These peptides will have no free N- or C-termini.

Accordingly in this aspect of the present invention there is provided a cyclised conotoxin peptide comprising a linear conotoxin peptide and a peptide linker, wherein the N- and C-termini of the linear peptide are linked via the peptide linker to form an amide cyclised peptide backbone.

No examples of cyclic conotoxins have been previously described in the literature, but it is in principle possible to make molecules which have a cyclic backbone, part of which incorporates the natural sequence and disulfide bond connections of linear conotoxins.

Cyclisation may also be achieved using other linking moieties , such as those including organic linkers, non-native peptide bonds such as thio-ether linkages and side-chain to N or C-termini cyclisation.

Considerable variation in the peptide sequence of the linking moiety is possible. Since this linking region does not bind to the primary active site of the conotoxin it can be modified to alter physiochemical properties, and potentially reduce side effects of the conotoxins.

In linking the N- and C-termini of the conotoxin it may in some cases be necessary or desirable to remove one or more of the N- or C-termini residues. Such modification of the linear conotoxin sequence is within the scope of the present invention.

The linking moiety will necessarily be of sufficient length to span the distance between the N- and C-termini of the conotoxin peptide. In the case of peptide linkers the length will generally be in the order of 2 to 15 amino acids. In some cases longer or shorter peptide linkers may be required.

Examples of possible peptide linkers include:

TRNGLPG	SEQ ID NO. 1
TRNG	SEQ ID NO. 2
TRGGLPV	SEQ ID NO. 3
TNG	SEQ ID NO. 4

It is possible, according to the present invention, to modify or potentiate the activity of a conotoxin peptide by selection of a particular size and/or type of peptide linker. Small changes in the conformation of the conotoxin caused by the introduction of a linking group can alter the binding affinities of the peptides for their particular binding sites. Conversely, where the activity is to be as close to the activity of the parent conotoxin peptide as possible, a linker will be selected which minimises any change in conformation.

There are several ways in which cyclic conotoxins may be synthesised. These include the following:

1. Cyclisation of the reduced peptide followed by oxidation to form the required disulfide bonds.

In this approach an extended linear peptide is first synthesised "on resin" using solid phase peptide synthesis methods. This extended linear peptide comprises the native sequence starting at a cysteine residue at, or closest to, the N-terminus and a C-terminal extension which comprises the new linking moiety. The solid phase synthesis actually starts in the reverse order- ie at the C-terminus of the extended linear peptide. Following cleavage from the resin, the extended conotoxin is cyclised to a thioester intermediate which subsequently rearranges to an amide-cyclised peptide. This reduced peptide is then oxidised to form the disulfide bonds. A schematic diagram of the reaction involved in the cyclisation is shown in Figure 2. The linear peptide is cleaved from the resin with the linker to the resin (R) still attached. R corresponds to the linker between the peptide and the resin and is different from the linking moiety used in the cyclisation. The first reaction involves the formation of a thioester between

the thiol of the N-terminal cysteine and the carboxy terminus. This then undergoes an S, N acyl migration to form the cyclic peptide with a native peptide bond.

2. Oxidation of the reduced linear peptide, followed by cyclisation.

In this approach an extended peptide is assembled using solid phase peptide synthesis. The extended linear peptide comprises the native conotoxin sequence with extra residues added at the N- and/or C-termini. The (new) N and C termini should preferably be glycine residues. The peptide is folded, and in the case of the conotoxin-like peptides, the termini of the folded molecule are generally close together in space. This facilitates the cyclisation of the peptide in solution using standard chemistry. Complications may occur when large numbers of lysine, glutamic acid or aspartic acid residues are present in the sequence and method 1 is then preferable.

3. Ligation of a linker onto an existing conotoxin, followed by cyclisation.

In this method the starting material is a mature conotoxin. A peptide linker is synthesised and ligated with the conotoxin using published procedures for the ligation of peptides. The extended peptide is then cyclised and oxidised.

Accordingly in a further aspect of the invention there is provided a process for preparing a cyclic conotoxin comprising:

A (i) synthesising an extended linear conotoxin peptide on a solid phase support, said extended linear conotoxin peptide comprising a linear conotoxin peptide having a linker moiety attached to at least one end thereof,
(ii) cleaving said extended linear peptide from the support
(iii) cyclising said extended linear conotoxin peptide, and
(iv) oxidising said cyclised peptide to form disulphide bonds, or

B (i) synthesising an extended linear conotoxin peptide on a solid phase support, said extended linear conotoxin peptide comprising a linear conotoxin peptide having a linker moiety attached to at least one end thereof,
(ii) cleaving said extended linear peptide from the solid support

(iii) subjecting said extended peptide to conditions such that the peptide folds and forms the required disulphide bonds, and
(iv) cyclising the folded peptide, or

C (i) reacting a conotoxin peptide with a linker moiety to form an extended linear conotoxin peptide having said linker moiety attached to one end thereof, and
(ii) cyclising said extended peptide and oxidising to form disulphide bonds, if required.

In the process described above the steps can be performed in any order, provided the product is a cyclic conotoxin having the required disulphide bonds. For example, in process A the cleavage and cyclisation steps may be performed simultaneously or in either order. Similarly in process B the cyclisation and folding steps could be performed simultaneously, or in either order.

It is also possible to form the disulphide bonds selectively using protecting groups on the cysteine residues. Selective protection of the cysteine residues in this way allows the production of a particular disulphide bond pattern. Examples of groups capable of protecting cysteine residues include acetamidomethyl (Acm), 4-methylbenzyl (MeBzl) and 4-methoxybenzyl (Mob).

Also, in view of the cyclic nature of the final products, synthetic procedures may involve cyclic permutation of the above procedures. For example, the designs of the extended linear peptide for α-conotoxins could commence by adding a linker to the C-terminal residue of the α-conotoxin, cyclically permuting the N-terminal residue(s) to the C-terminal, to provide an N-terminal cysteine, and cyclising as described.

Some examples of linear conotoxins which are currently known and to which the cyclisation approach can be applied are listed in Table 1.

Table 1. Amino acid sequences of selected known conotoxins.

Conotoxin	Sequence
Omega conotoxins	
MVIIA	CKGKGAKCSRLMYDCCTGSCRS--GKC
MVIIC	CKGKGAOCRKTMYDCCSGSCGRR-GKC
GVIA	CKSOGSSCSOTSYNCCR-SCNOYTKRCY
SVIA	CRSSGSOCGVTSI-CCGR-CYR--GKCT
SVIB	CKLKGQSCRKTSYDCCSGSCGRS-GKC
GVIIA	CKSOGTOCSRGMRDCCTS-CLLYSNKCRRY
GVIIB	CKSOGTOCSRGMRDCCTS-CLSYSNKCRRY
TVIA	CLSOGSSCSOTSYNCCRS-CNOYSRKCR
Kappa conotoxin	
PVIIA	CRIONQKCFQHLDDCCSRKCNRFNKCV
Alpha conotoxins	
GI	ECCNPA-CGRHYS--C
IMI	GCCSDPRCAWR----C
PNIA	GCCSLPPCAANNPDYC
PNIB	GCCSLPPCALSNPDYC
SII	GCCCNPACGPNYG--CGTSCS
MII	GCCSNPBCHLEHSNLC
Mu conotoxins	
GIIIA	-RDCCTOOKKCKDRQCKOQRCCA
GIIIB	-RDCCTOORKCKDRRCKOMKCCA
GIIIC	-RDCCTOOKKCKDRRCKOLKCCA
PIIIA	ZRLCCGFOKSCRSRQCKOHRCC
GS	ACSGRGSRCPPQCCMGLRCGRGNPQKCIGAHEDV

The term "derivative" as used herein in connection with naturally occurring conotoxin peptides, such as MVIIA, refers to a peptide which differs from the naturally occurring

peptides by one or more amino acid deletions, additions, substitutions, or side-chain modifications.

Substitutions encompass amino acid alterations in which an amino acid is replaced with a different naturally-occurring or a non-conventional amino acid residue. Such substitutions may be classified as "conservative", in which case an amino acid residue contained in a polypeptide is replaced with another naturally-occurring amino acid of similar character either in relation to polarity, side chain functionality, or size, for example Ser↔Thr↔Pro↔Hyp↔Gly↔Ala, Val↔Ile↔Leu, His↔Lys↔Arg, Asn↔Gln↔Asp↔Glu or Phe↔Trp↔Tyr. It is to be understood that some non-conventional amino acids may also be suitable replacements for the naturally occurring amino acids. For example ornithine, homoarginine and dimethyllysine are related to His, Arg and Lys.

Substitutions encompassed by the present invention may also be "non-conservative", in which an amino acid residue which is present in a polypeptide is substituted with an amino acid having different properties, such as a naturally-occurring amino acid from a different group (eg. substituting a charged or hydrophobic amino acid with alanine), or alternatively, in which a naturally-occurring amino acid is substituted with a non-conventional amino acid.

Amino acid substitutions are typically of single residues, but may be of multiple residues, either clustered or dispersed.

Preferably, amino acid substitutions are conservative.

Additions encompass the addition of one or more naturally occurring or non-conventional amino acid residues. Deletion encompasses the deletion of one or more amino acid residues.

As stated above the present invention includes peptides in which one or more of the amino

acids has undergone sidechain modifications. Examples of side chain modifications contemplated by the present invention include modifications of amino groups such as by reductive alkylation by reaction with an aldehyde followed by reduction with $NaBH_4$; amidination with methylacetimidate; acylation with acetic anhydride; carbamoylation of amino groups with cyanate; trinitrobenzylation of amino groups with 2, 4, 6-trinitrobenzene sulphonic acid (TNBS); acylation of amino groups with succinic anhydride and tetrahydrophthalic anhydride; and pyridoxylation of lysine with pyridoxal-5-phosphate followed by reduction with $NaBH_4$.

The guanidine group of arginine residues may be modified by the formation of heterocyclic condensation products with reagents such as 2,3-butanedione, phenylglyoxal and glyoxal.

The carboxyl group may be modified by carbodiimide activation *via* O-acylisourea formation followed by subsequent derivitisation, for example, to a corresponding amide.

Sulphydryl groups may be modified by methods such as carboxymethylation with iodoacetic acid or iodoacetamide; performic acid oxidation to cysteic acid; formation of mixed disulphides with other thiol compounds; reaction with maleimide, maleic anhydride or other substituted maleimide; formation of mercurial derivatives using 4-chloromercuribenzoate, 4-chloromercuriphenylsulphonic acid, phenylmercury chloride, 2-chloromercuri-4-nitrophenol and other mercurials; carbamoylation with cyanate at alkaline pH. Any modification of cysteine residues must not affect the ability of the peptide to form the necessary disulphide bonds. It is also possible to replace the sulphydryl groups of cysteine with selenium equivalents such that the peptide forms a diselenium bond in place of one or more of the disulphide bonds.

Tryptophan residues may be modified by, for example, oxidation with N-bromosuccinimide or alkylation of the indole ring with 2-hydroxy-5-nitrobenzyl bromide or sulphenyl halides. Tyrosine residues on the other hand, may be altered by nitration with tetranitromethane to form a 3-nitrotyrosine derivative.

Modification of the imidazole ring of a histidine residue may be accomplished by alkylation with iodoacetic acid derivatives or N-carbethoxylation with diethylpyrocarbonate.

Proline residues may be modified by, for example, hydroxylation in the 4-position.

A list of some amino acids having modified side chains and other unnatural amino acids is shown in Table 2.

TABLE 2

Non-conventional amino acid	Code	Non-conventional amino acid	Code
α-aminobutyric acid	Abu	L-N-methylalanine	Nmala
α-amino-α-methylbutyrate	Mgabu	L-N-methylarginine	Nmarg
aminocyclopropane-carboxylate	Cpro	L-N-methylasparagine	Nmasn
		L-N-methylaspartic acid	Nmasp
aminoisobutyric acid	Aib	L-N-methylcysteine	Nmcys
aminonorbornyl-carboxylate	Norb	L-N-methylglutamine	Nmgln
		L-N-methylglutamic acid	Nmglu
cyclohexylalanine	Chexa	L-N-methylhistidine	Nmhis
cyclopentylalanine	Cpen	L-N-methylisolleucine	Nmile
D-alanine	Dal	L-N-methylleucine	Nmleu
D-arginine	Darg	L-N-methyllysine	Nmlys
D-aspartic acid	Dasp	L-N-methylmethionine	Nmmet
D-cysteine	Dcys	L-N-methylnorleucine	Nmnle
D-glutamine	Dgln	L-N-methylnorvaline	Nmnva
D-glutamic acid	Dglu	L-N-methylornithine	Nmorn
D-histidine	Dhis	L-N-methylphenylalanine	Nmphe
D-isoleucine	Dile	L-N-methylproline	Nmpro
D-leucine	Dleu	L-N-methylserine	Nmser

D-lysine	Dlys	L-N-methylthreonine	Nmthr
D-methionine	Dmet	L-N-methyltryptophan	Nmtrp
D-ornithine	Dorn	L-N-methyltyrosine	Nmtyr
D-phenylalanine	Dphe	L-N-methylvaline	Nmval
D-proline	Dpro	L-N-methylethylglycine	Nmetg
D-serine	Dser	L-N-methyl-t-butylglycine	Nmtbug
D-threonine	Dthr	L-norleucine	Nle
D-tryptophan	Dtrp	L-norvaline	Nva
D-tyrosine	Dtyr	α-methyl-aminoisobutyrate	Maib
D-valine	Dval	α-methyl-γ-aminobutyrate	Mgabu
D-α-methylalanine	Dmala	α-methylcyclohexylalanine	Mchexa
D-α-methylarginine	Dmarg	α-methylcylcopentylalanine	Mcpen
D-α-methylasparagine	Dmasn	α-methyl-α-napthylalanine	Manap
D-α-methylaspartate	Dmasp	α-methylpenicillamine	Mpen
D-α-methylcysteine	Dmcys	N-(4-aminobutyl)glycine	Nglu
D-α-methylglutamine	Dmgln	N-(2-aminoethyl)glycine	Naeg
D-α-methylhistidine	Dmhis	N-(3-aminopropyl)glycine	Norn
D-α-methylisoleucine	Dmile	N-amino-α-methylbutyrate	Nmaabu
D-α-methylleucine	Dmleu	α-napthylalanine	Anap
D-α-methyllysine	Dmlys	N-benzylglycine	Nphe
D-α-methylmethionine	Dmmet	N-(2-carbamylethyl)glycine	Ngln
D-α-methylornithine	Dmorn	N-(carbamylmethyl)glycine	Nasn
D-α-methylphenylalanine	Dmphe	N-(2-carboxyethyl)glycine	Nglu
D-α-methylproline	Dmpro	N-(carboxymethyl)glycine	Nasp
D-α-methylserine	Dmser	N-cyclobutylglycine	Ncbut
D-α-methylthreonine	Dmthr	N-cycloheptylglycine	Nchep
D-α-methyltryptophan	Dmtrp	N-cyclohexylglycine	Nchex
D-α-methyltyrosine	Dmty	N-cyclodecylglycine	Ncdec
D-α-methylvaline	Dmval	N-cylcododecylglycine	Ncdod
D-N-methylalanine	Dnmala	N-cyclooctylglycine	Ncoct
D-N-methylarginine	Dnmarg	N-cyclopropylglycine	Ncpro

	D-N-methylasparagine	Dnmasn	N-cycloundecylglycine	Ncund
	D-N-methylaspartate	Dnmasp	N-(2,2-diphenylethyl)glycine	Nbhm
	D-N-methylcysteine	Dnmcys	N-(3,3-diphenylpropyl)glycine	Nbhe
	D-N-methylglutamine	Dnmgln	N-(3-guanidinopropyl)glycine	Narg
	D-N-methylglutamate	Dnmglu	N-(1-hydroxyethyl)glycine	Nthr
	D-N-methylhistidine	Dnmhis	N-(hydroxyethyl))glycine	Nser
	D-N-methylisoleucine	Dnmile	N-(imidazolylethyl))glycine	Nhis
	D-N-methylleucine	Dnmleu	N-(3-indolylyethyl)glycine	Nhtrp
	D-N-methyllysine	Dnmlys	N-methyl-γ-aminobutyrate	Nmgabu
	N-methylcyclohexylalanine	Nmchexa	D-N-methylmethionine	Dnmmet
	D-N-methylornithine	Dnmorn	N-methylcyclopentylalanine	Nmcpen
	N-methylglycine	Nala	D-N-methylphenylalanine	Dnmphe
	N-methylaminoisobutyrate	Nmaib	D-N-methylproline	Dnmpro
	N-(1-methylpropyl)glycine	Nile	D-N-methylserine	Dnmser
	N-(2-methylpropyl)glycine	Nleu	D-N-methylthreonine	Dnmthr
	D-N-methyltryptophan	Dnmtrp	N-(1-methylethyl)glycine	Nval
	D-N-methyltyrosine	Dnmtyr	N-methyla-napthylalanine	Nmanap
	D-N-methylvaline	Dnmval	N-methylpenicillamine	Nmpen
	γ-aminobutyric acid	Gabu	N-(*p*-hydroxyphenyl)glycine	Nhtyr
	L-*t*-butylglycine	Tbug	N-(thiomethyl)glycine	Ncys
	L-ethylglycine	Etg	penicillamine	Pen
	L-homophenylalanine	Hphe	L-α-methylalanine	Mala
	L-α-methylarginine	Marg	L-α-methylasparagine	Masn
	L-α-methylaspartate	Masp	L-α-methyl-*t*-butylglycine	Mtbug
	L-α-methylcysteine	Mcys	L-methylethylglycine	Metg
	L-α-methylglutamine	Mgln	L-α-methylglutamate	Mglu
	L-α-methylhistidine	Mhis	L-α-methylhomophenylalanine	Mhphe
	L-α-methylisoleucine	Mile	N-(2-methylthioethyl)glycine	Nmet
	L-α-methylleucine	Mleu	L-α-methyllysine	Mlys
	L-α-methylmethionine	Mmet	L-α-methylnorleucine	Mnle
	L-α-methylnorvaline	Mnva	L-α-methylornithine	Morn

L-α-methylphenylalanine	Mphe	L-α-methylproline	Mpro
L-α-methylserine	Mser	L-α-methylthreonine	Mthr
L-α-methyltryptophan	Mtrp	L-α-methyltyrosine	Mtyr
L-α-methylvaline	Mval	L-N-methylhomophenylalanine	Nmhphe
N-(N-(2,2-diphenylethyl) carbamylmethyl)glycine	Nnbhm	N-(N-(3,3-diphenylpropyl) carbamylmethyl)glycine	Nnbhe
1-carboxy-1-(2,2-diphenyl-ethylamino)cyclopropane	Nmbc	L-O-methyl-serine	Omser
		L-O-methyl homoserine	Omhser

These types of modifications may be important to stabilise the peptide if administered to an individual or for use as a diagnostic reagent.

Other derivatives contemplated by the present invention include a range of glycosylation variants from a completely unglycosylatecd molecule to a modified glycosylated molecule. Altered glycosylation patterns may result from expression of recombinant molecules in different host cells.

Preferably cyclic conotoxin peptides will retain the Cys residues and characteristic disulphide bonding pattern. Derivatives may include additional Cys residues provided they are protected during formation of the disulphide bonds.

Preferably the conotoxin peptides according to the invention have 12 to 40 amino acids, more preferably 15 to 30.

Naturally occurring conotoxins are widely used as neuropharmacological probes. They bind very tightly and highly selectivity to ion channel receptors. In these applications they are incubated with a relevant tissue preparation and their binding, or biological effects are measured. Their actions will be reduced or destroyed if they are metabolized by endogenous enzymes. Optimum performance of pharmacological probes thus requires resistance to enzymatic or chemical breakdown. Since the cyclic conotoxin peptides

possess the desirable properties described above they may be better pharmacological probes than naturally occurring conotoxin peptides in some cases.

Still another aspect of the present invention is directed to antibodies to the cyclic peptides according to the invention. Such antibodies may be monoclonal or polyclonal and may be selected from naturally occurring antibodies to the peptides or may be specifically raised to the peptides using standard techniques. In the case of the latter, the peptides may first need to be associated with a carrier molecule. The antibodies of the present invention are particularly useful as therapeutic or diagnostic agents.

In this regard, specific antibodies can be used to screen for the peptides according to the invention. Techniques for such assays are well known in the art and include, for example, sandwich assays and ELISA. Knowledge of peptide levels may be important for monitoring certain therapeutic protocols.

The cyclic conotoxin peptides according to the present invention are useful as therapeutic agents.

Accordingly the present invention provides a method for the treatment or prophylaxis of conditions or diseases in mammals, preferably humans, including the step of administering a cyclic conotoxin peptide.

In particular omega-conotoxins which block N-type calcium channels may be useful in the treatment of neurological disorders such as acute and chronic pain, stroke, traumatic brain injury, migraine, epilepsy, Parkinson's disease, Alzheimer's disease, multiple schlerosis, and depression. The α-conotoxins bind to nicotinic acetylcholine receptors (nAChRs). Such receptors have been implicated in the pathophysiology of several neuropsychiatric disorders including schizophrenia, Alzheimer's disease, Parkinson's disease and Tourette's syndrome and thus the α-conotoxins have potential therapeutic indications for these diseases. The μ-conotoxins target sodium channels. Those μ-conotoxins that interact with neuronal channels (eg PIIIA) have potential therapeutical

applications in the treatment of chronic and neuropathic pain.

Assays useful for assessing compounds with the above mentioned activities may be *in vitro* or *in vivo* and are known to those skilled in the art. For example, assays useful for assessing activity at N-type calcium channels include those described or referenced in WO91/07980, WO93/13128, US 5,824,645, WO97/04797, Drugs of the Future (1994 and 1998), Drug Data Report (1993), or Heading (1999). The cyclic peptides according to the invention, or labelled derivatives thereof, may also be useful in such assays.

Preferably the mammal is in need of such treatment although the peptide may be administered in a prophylactic sense.

The invention also provides a composition comprising a cyclic conotoxin peptide, and a pharmaceutically acceptable carrier or diluent.

Preferably the composition is in the form of a pharmaceutical composition.

There is also provided the use of a cyclic conotoxin peptide in the manufacture of a medicament for the treatment or prophylaxis of diseases or conditions of mammals, preferably of humans.

As will be readily appreciated by those skilled in the art, the route of administration and the nature of the pharmaceutically acceptable carrier will depend on the nature of the condition and the mammal to be treated. It is believed that the choice of a particular carrier or delivery system, and route of administration could be readily determined by a person skilled in the art. In the preparation of any formulation containing the peptide actives care should be taken to ensure that the activity of the peptide is not destroyed in the process and that the peptide is able to reach its site of action without being destroyed. In some circumstances it may be necessary to protect the peptide by means known in the art, such as, for example, micro encapsulation. Similarly the route of administration chosen should be such that the peptide reaches its site of action. In view of the improved

stability of the cyclic peptides relative to their "linear" counterparts a wider range of formulation types and routes of administration is available. Known conotoxins can generally only be administered successfully intrathecally which means that the patient must be hospitalised. Administration of the cyclic peptides according to the present invention is not subject to the same restriction.

The pharmaceutical forms suitable for injectable use include sterile injectable solutions or dispersions, and sterile powders for the extemporaneous preparation of sterile injectable solutions. They should be stable under the conditions of manufacture and storage and may be preserved against the contaminating action of microorganisms such as bacteria or fungi . The solvent or dispersion medium for the injectable solution or dispersion may contain any of the conventional solvent or carrier systems for peptide actives, and may contain, for example, water, ethanol, polyol (for example, glycerol, propylene glycol and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be brought about where necessary by the inclusion of various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal and the like. In many cases, it will be preferable to include agents to adjust osmolality, for example, sugars or sodium chloride. Preferably, the formulation for injection will be isotonic with blood. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin. Pharmaceutical forms suitable for injectable use may be delivered by any appropriate route including intravenous, intramuscular, intracerebral, intrathecal injection or infusion.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains

the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from previously sterile-filtered solution thereof.

When the active ingredient is suitably protected it may be orally administered, for example, with an inert diluent or with an assimilable edible carrier, or it may be enclosed in hard or soft shell gelatin capsule, or it may be compressed into tablets, or it may be incorporated directly with the food of the diet. For oral therapeutic administration, the active compound may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations preferably contain at least 1% by weight of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 5 to about 80% of the weight of the unit. The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained.

The tablets, troches, pills, capsules and the like may also contain the components as listed hereafter: A binder such as gum, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such a sucrose, lactose or saccharin may be added or a flavouring agent such as peppermint, oil of wintergreen, or cherry flavouring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. A syrup or elixir may contain the active compound, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavouring such as cherry or orange flavour. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active

compound(s) may be incorporated into sustained-release preparations and formulations.

The present invention also extends to any other forms suitable for administration, for example topical application such as creams, lotions and gels, or compositions suitable for inhalation or intranasal delivery, for example solutions or dry powders.

Parenteral dosage forms are preferred, including those suitable for intravenous, intrathecal, or intracerebral delivery.

Pharmaceutically acceptable carriers and/or diluents include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the novel dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active material and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active material for the treatment of disease in living subjects having a diseased condition in which bodily health is impaired as herein disclosed in detail.

The principal active ingredient is compounded for convenient and effective administration in effective amounts with a suitable pharmaceutically acceptable carrier in dosage unit form. A unit dosage form can, for example, contain the principal active compound in amounts ranging from 0.25 μg to about 2000 mg. Expressed in proportions, the active

compound is generally present in from about 0.25 μg to about 2000 mg/ml of carrier. In the case of compositions containing supplementary active ingredients, the dosages are determined by reference to the usual dose and manner of administration of the said ingredients.

The invention will now be described with reference to the accompanying examples and figures which describe the production of some cyclic conotoxin peptides and their biological activity and illustrate the structures of some linear conotoxin peptides which may be subjected to cyclisation. However, it is to be understood that the particularity of the following description is not to supersede the generality of the preceding description of the invention.

Referring to the figures:

Figure 1 is a representation of the three-dimensional structures of the conotoxins PVIIA, MVIIA and GS. The structures were determined by NMR spectroscopy. The backbone atoms are displayed as lines and the disulfide bonds are highlighted as balls and sticks. All three conotoxins, although from different classes and hence having different activities, have similar structures which contain a cystine knot motif.

Figure 2 is a scheme for peptide cyclisation via a C-terminal thioester. The free sulfur of an N-terminal cysteine interacts with the C-terminal thioester to form an intermediate which undergoes an S,N, acyl migration to form a cyclic peptide with a native peptide bond.

Figure 3 is a representation of the three-dimensional structures of the conotoxins PVIIA, MVIIA, SVIB, GI and IMI. The structures were determined by NMR spectroscopy. The backbone atoms are shown as lines and the N- and C-termini are connected by a dotted line which has the intervening distance shown above. The α-conotoxins, GI and IMI, have slightly closer termini than the conotoxins shown in the top of the figure, which suggests cyclization would be more feasible for this class of conotoxins and may

even occur more readily than that shown for MVIIA.

EXAMPLES

Example 1

A cyclic analogue of MVIIA (cyclo-MVIIA 1) has been synthesised with the sequence :-

CKGKGAKCSRLMYDCCTGSCRSGKCTRNGLPG SEQ ID NO. 5

The residues in bold represent the sequence of MVIIA. Those not in bold are the linking moiety (TRNGLPG). A thioester method has been used in the synthesis of this peptide which was performed on a Gly PAM resin. A - SCH_2-CH_2CO- linker was attached to the Gly-PAM resin by treating the resin with bromopropanoic acid for 30 minutes, washing with DMF and then treating the resin with 10% thioacetic acid, 10% DIEA in DMF for 2 x 20 minutes. The resin was again washed with DMF and treated with 10% β-mercaptoethanol, 10% DIEA in DMF for 2 x 20 minutes. After a final wash with DMF, the first residue, Boc-glycine, was coupled to the resin using HBTU and DIEA. The remainder of the peptide was assembled by manual synthesis using HBTU with *in situ* neutralisation (Schnölzer, M. *et al.*, 1992).

The linker is not stable under basic conditions, thus the formyl group was not removed from the tryptophan with ethanolamine prior to HF cleavage. Cresol (800 µL) and thiocresol (200 µL) were used as scavengers during the HF cleavage which was carried out for 2 hours at -2 to 0 °C. The crude, reduced peptide was purified using preparative reverse-phase HPLC on a Vydac C18 column. Gradients of 0.1 % aqueous TFA and 90% acetonitrile/0.09% TFA were employed with a flow rate of 8 mL/min and the eluant monitored at 230 nm. The reduced peptide was cyclised in 0.1 M sodium phosphate (pH 7.4), with a 6 fold excess of TCEP at room temperature for 30 minutes. All linear material was cyclised within this time as judged by analytical reverse phase HPLC and mass spectrometry. Mass analysis was performed on a Sciex (Thornhill, Ontario) triple quadrupole mass spectrometer using electrospray sample ionization. Cyclo-MVIIA 1 was oxidized at a concentration of 0.5 mg/ml in 2M $(NH_4)_2SO_4$, 0.1 M NH_4OAc (pH8) and 1mM reduced glutathione at 4°C for 24 hours. The product was purified using reverse phase preparative HPLC.

Example 2

A slightly smaller cyclic analogue of MVIIA (cyclo-MVIIA 2) has been synthesised with the sequence:-

CKGKGAKCSRLMYDCCTGSCRSGKCTRNG SEQ ID NO. 6

Once again the bold residues correspond to the sequence of MVIIA, (all except TRNG). This peptide was synthesised using the procedures outlined in Example 1. Following cyclisation, cyclo-MVIIA 2 was oxidised at a concentration of 0.5 mg/mL in 2 M $(NH_4)_2SO_4$, 0.1 M NH_4OAc (pH 8) and 1 mM reduced glutathione at 4 °C for 24 hours. Three major components were present in the oxidation and were all purified using a semi-preparative C18 column (3mL/min) with monitoring at 230 nm. The three components represent cyclic fully disulfide bonded forms of cyclo-MVIIA 2.

Example 3

a) Antagonists specific to N-type voltage-sensitive calcium channels are being used as leads in drug development. Examples of these are ω-conotoxins GVIA and MVIIA. An assay has previously been established to determine the ability of a compound to displace ^{125}I-GVIA from receptors in rat membrane. Rat membrane was prepared according to the procedure of Wagner *et al.* 1988. Rats were sacrificed by cervical dislocation and their brains removed and immediately frozen in liquid nitrogen. Frozen brains were stored at -78 °C until required. Three brains (wet weight , 6.25 g) were thawed (50 mM HEPES, pH 7.4) and homogenised with ultraturrex (IKA, 170 Watt) in 125 mls 50 mM HEPES pH 7.4. Homogenised brain was centrifuged at 16000rpm (35000g) for 20min at 4 °C and the supernatant discarded. The pellet was resuspended by further homogenisation in 50mM HEPES, pH 7.4, 10 mM EDTA and incubated at 4 °C for 30 min. Centrifugation was repeated as above and the supernatant discarded. The pellet was resuspended in 125ml 50mM HEPES, pH 7.4 (1:20 dilution) and stored at -78 °C.

^{125}I-[Tyr22]GVIA was prepared according to the procedure of Cruz and Olivera (1986) and isolated by reverse-phase HPLC on an analytical Vydac C18 column. The column was equilibrated in buffer A (H_2O, 0.1% TFA) followed by a linear gradient to 67% buffer B (90% acetonitrile, 10% H_2O and 0.09% TFA) in 100 min. Peaks were detected at 214 nm and the flow rate was 1 ml/min. The radiolabeled peaks were counted using a gamma counter and stored at 4 °C.

Assays were performed in 12 x 75 mm borasilicate culture tubes at room temperature and incubated for 1hr. Each tube contained 100μl each of test solution, iodinated ligand (7 fmol) and rat membrane (16 μg) added in this order. The assay buffer contained 20mM HEPES pH7.2, 75 mM NaCl, 0.1 mM EDTA, 0.1 mM EGTA, 0.1% BSA and protease inhibitors, 2 mM leupeptin and 0.5U aprotinin. The nonspecific binding was determined in the presence of 17nM GVIA. Assays were terminated by vacuum filtration on a Millipore manifold filtration system using glass fibre filters (Whatman GFB) presoaked in 0.6% polyethylenimine. Each tube was washed 3 times with 3ml ice-cold wash buffer (20mM HEPES pH7.2, 125mM NaCl and 0.1% BSA). Filters were counted on a gamma counter. Graphpad Prism was used to generate binding curves and calculate EC_{50} values. The EC_{50} values are a measure of the ability of a compound to displace ^{125}I-GVIA; the EC_{50} for MVIIA is 4.4 x 10^{-11} M. Fractions isolated from oxidation of the cysteine residues in cyclic, reduced cyclo-MVIIA 1 were tested in this assay. As expected, not all disulfide isomers had the same level of activity. The most active isomer exhibited an EC50 of 8.5 x 10-8 M. The three oxidized, cyclic forms of cyclo-MVIIA 2 were also tested in this assay and the most active isomer exhibited an EC50 of 5 x 10-10 M. As expected, not all disulfide isomers had the same level of activity.

b) To test the specificity of the cyclic conotoxin derivatives for the N-type Ca channel relative to P/Q type channels additional binding studies were done using ^{125}I-MVIIC as the displaced ligand. This binds selectively to P/Q type Ca channels.

The assay was carried out as described in Example 3a, except that ^{125}I-MVIIC (selective for P/Q-type) channels was used as the displaced ligand rather than ^{125}I-GVIIA (selective for N-type channels). The ^{125}I-MVIIC was prepared and purified as described in Nielsen et al, 1999.

The most active form of cyclo-MVIIA 2 did not show any ability to displace ^{125}I-MVIIC when administered at concentrations up to 630 nM. When combined with the data described above for displacement of ^{125}I-GVIIA from N-type channels, this demonstrates selectivity for the N-type channel over the P/Q-type channel.

Example 4

The three-dimensional structures of several conotoxin peptides have been determined by NMR spectroscopy to confirm the feasibility of making cyclic conotoxins which do not significantly alter the conformation of most parts of the conotoxin molecules. A comparison of five conotoxin structures determined by NMR is presented in figure 3.

Only the backbone atoms are displayed and the amino and carboxy termini are labelled as N and C respectively. The distances in angstroms between the termini have been measured and are also marked on the diagram. The three structures in the top half of the diagram represent PVIIA (Scanlon *et al.*, 1997), MVIIA (Nielsen *et al.*, 1996) and SVIB (Nielsen *et al.*, 1996). It is clear that in all three peptides the overall structure is very similar, as is the distance between the termini. MVIIA and SVIB are both classed as omega conotoxins and have some sequence homology (Table 1), however PVIIA belongs to the kappa class and has little sequence homology to MVIIA and SVIB except for the conserved cysteine residues. It has now been shown that MVIIA can by cyclised and still retain a high level of activity (Examples 1-3). Given the structural similarity between the peptides mentioned above, cyclisation is feasible for other conotoxins, such as PVIIA and SVIB.

The alpha conotoxins have a different structure than the previously mentioned peptides,

however the termini are still close, as shown for GI (Gehrmann *et al.*, 1998) and IMI (unpublished data) above. The close proximity of the termini suggests cyclisation can be achieved without significantly affecting the biological activity. Thus, the concept of cyclising conotoxins is applicable not only to omega conotoxins but to peptides from other classes of conotoxins, including alpha and kappa, and extends to all conotoxins which have termini located close together, especially those within a distance of approximately 13 Å (i.e. the distance present in MVIIA).

In the case of mu-conotoxins the termini are further apart in general, but cyclisation is readily possible using longer peptide sequences as linkers. In the case of Na-channel conotoxins like GS the peptide contains a C-terminal extension beyond the final cysteine residue that may form part of the cyclising linker.

Example 5

To exemplify the principles involved in synthetic method 2 described above an analogue of MVIIA has been synthesised using solid phase peptide synthesis with Boc chemistry. The synthesised peptide has the sequence:

GLPV**CKGKGAKC SRLMYDCC TGSCRSGKC**TRG SEQ ID NO. 7

The peptide has both an N(GLPU) and C(TRG) terminal extension and the remaining residues (in bold) represent MVIIA. The reduced peptide was purified using the conditions given in Examples 1 and 2. Oxidation was achieved using 0.1 M ammonium acetate, 2M ammonium sulfate, pH 7.7, 1mM reduced glutathione and the reaction left at 4°C for two days. The oxidised peptide was purified and the activity tested as in Example 3. An EC_{50} of 1.081×10^{-9} M was found for this analogue, illustrating that extending the N and C termini of the peptide, as may be necessary prior to cyclisation, does not eliminate activity.

Example 6

A cyclic α-conotoxin is prepared based on the sequence of α-contoxin MII. The linear precursor for this synthesis is designed by first adding a linker moiety to the native sequence as shown below. The residues in bold correspond to the native sequence of MII and the non-bold residues are the linker moiety (TNG).

GCCSNPVCHLEHSNLCTNG SEQ ID NO. 8

A cyclically permuted derivative of this sequence is then designed by moving the N-terminal glycine residue to the C-terminus to produce the sequence:

CCSNPVCHLEHSNLCTNGG SEQ ID NO. 9

Thi peptide is synthesised using the thioester method described above in which the C-terminal glycine is attached to a Gly PAM resin via a $-SCH_2CH_2CO-$ linker. The linker is attached to the Gly PAM resin by treating the resin with bromopropanoic acid for 30 minutes, washing with DMF and then treating the resin with 10% thioacetic acid, 10% DIEA in DMF for 2 x 20 minutes. The resin is washed again with DMF and treated with 10% β-mercaptoethanol, 10% DIEA in DMF for 2 x 20 minutes. After a final wash with DMF, the first (ie C-terminal) residue of the linear peptide sequence is coupled to the resin using HBTU and DIEA. The remainder of the peptide sequence is assembled by manual synthesis using HBTU with *in situ* neutralisation. Cleavage from the resin, cyclisation and oxidation is achieved using the methods described in Examples 1 and 2.

Example 7

The bioavailability of cyclic conotoxins is tested by either oral administration or intravenous administration into rats. Male Sprague-Dawley-derived rats (ca. 325g) are maintained on standard rat pellets until surgery, and are subsequently prepared, under isoflurane anaesthesia, with a catheter in the right external jugular vein. Rats are then

placed unrestrained n metabolism cages and allowed to recover prior to dosing. A 75 mm oral dosing (gavage) needle is used to dose conscious rats and the jugular catheter is used for iv dosing. Following dosing, plasma samples are taken out at time points between 0 and 180 min. A blood sample (ca. 500 mL) is withdrawn and centrifuged and then placed on ice until processing. The supernatant (200 mL) is transferred and HPLC grade acetonitrile (300 mL) added to precipitate proteins, however the test peptides remain in solution. The sample is then centrifuged and the supernatant transferred for further analysis. The supernatant is diluted with 0.1% TFA and injected on to an analytical reverse phase C18 column using gradients of 0.1% TFA/0.9% TFA in 90% acetonitrile:10% water. The eluent is monitored at 214 nm. This analysis allows calculation of a half-life for the peptide of interest.

Further studies are performed to give indications of stability of cyclic conotoxins in biological media and hence an indication of bioavailability. Biological media such as fetal calm serum and rat gastric juices are used. The cyclic conotoxin solution (10 mL 1mg/mL) is diluted with 0.1 M PBS pH 7.6 (~50 mL) and fetal calm serum (~50 mL) is added to the sample. The sample is then incubated at 37°C for ~1-5 hours. An aliquot (~40 mL) is removed and diluted with 0.1% TFA and injected on an analytical C18 reverse phase HPLC column with gradients of 0.1% TFA/0.9% TFA in acetonitrile. The sample is monitored at 214 nm. The stock peptide solution, appropriately diluted, is used as a control and allows the percentage breakdown at a particular timepoint to be calculated. A similar protocol is applied for rat gastric juices. However, the peptide is not diluted in buffer but incubated at 37°C for 1-5 hours and aliquots analyzed by reverse phase HPLC. Performing these studies on linear and cyclic conotoxins shows the greater stability of the cyclic conotoxins.

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Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps.

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

THE CLAIMS:

1. A cyclised conotoxin peptide.

2. A cyclised conotoxin peptide having an activity associated with the therapeutic treatment of mammals.

3. A cyclic conotoxin peptide which contains or consists of the sequence of amino acids present in a naturally occurring conotoxin peptide or derivative thereof.

4. A cyclic conotoxin peptide according to claim 3 wherein the naturally occurring conotoxin peptide is selected from MVIA, GVIA, SVIB, SVIA, TVIA, MVIIC, GVIIA, GVIIB, PVIIA, GS, GI, IMI, PNIA, PNIB, SII, MII, GIIIA, GIIIB, GIIIC and PIIIA.

5. A cyclic conotoxin peptide having three disulphide bonds in the form of a cysteine knot.

6. A cyclic conotoxin peptide comprising a linear conotoxin peptide and a peptide linker, wherein the N- and C- termini of the linear peptide are linked via the peptide linker to form an amide cyclised peptide backbone.

7. A cyclic conotoxin peptide according to claim 6 wherein the linear conotoxin peptide moiety is derived from a naturally occurring conotoxin peptide and retains the disulphide bond connectivity of the naturally occurring conotoxin peptide.

8. A cyclic conotoxin peptide according to claim 6 wherein the peptide linker is from 2 to 15 amino acids in length.

9. A cyclic conotoxin peptide according to claim 6 wherein the peptide linker is selected from the group consisting of:

TRNGLPG	SEQ ID NO. 1
TRNG	SEQ ID NO. 2
TRGGLPV	SEQ ID NO. 3
TNG	SEQ ID NO. 4

10. A cyclic conotoxin peptide selected from the group consisting of:

CKGKGAKCSRLMYDCCTGSCRSGKCTRNGLPG SEQ ID NO. 5

CKGKGAKCSRLMYDCCTGSCRSGKCTRNG SEQ ID NO. 6

GLPV**CKGKGAKCSRLMYDCCTGSCRSGKC**TRG SEQ ID NO. 7

GCCSNPVCHLEHSNLCTNG SEQ ID NO. 8

CCSNPVCHLEHSNLCTNG**G** SEQ ID NO. 9

11. A process for preparing a cyclic conotoxin comprising:
(i) synthesising an extended linear conotoxin peptide on a solid phase support, said extended linear conotoxin peptide comprising a linear conotoxin peptide having a linker moiety attached to at least one end thereof,
(ii) cleaving said extended linear peptide from the support
(iii) cyclising said extended linear conotoxin peptide, and
(iv) oxidising said cyclised peptide to form disulphide bonds.

12. A process for preparing a cyclic conotoxin comprising:
(i) synthesising an extended linear conotoxin peptide on a solid phase support, said

extended linear conotoxin peptide comprising a linear conotoxin peptide having a linker moiety attached to at least one end thereof,

(ii) cleaving said linear peptide from the solid support,

(iii) subjecting said extended peptide to conditions such that the peptide folds and forms the required disulphide bonds, and

(iv) cyclising the folded peptide.

13. A process for preparing a cyclic conotoxin comprising:

(i) reacting a conotoxin peptide with a linker moiety to form an extended linear conotoxin peptide having said linker moiety attached to one end thereof, and

(ii) cyclising said extended peptide and oxidising to form disulphide bonds, if required.

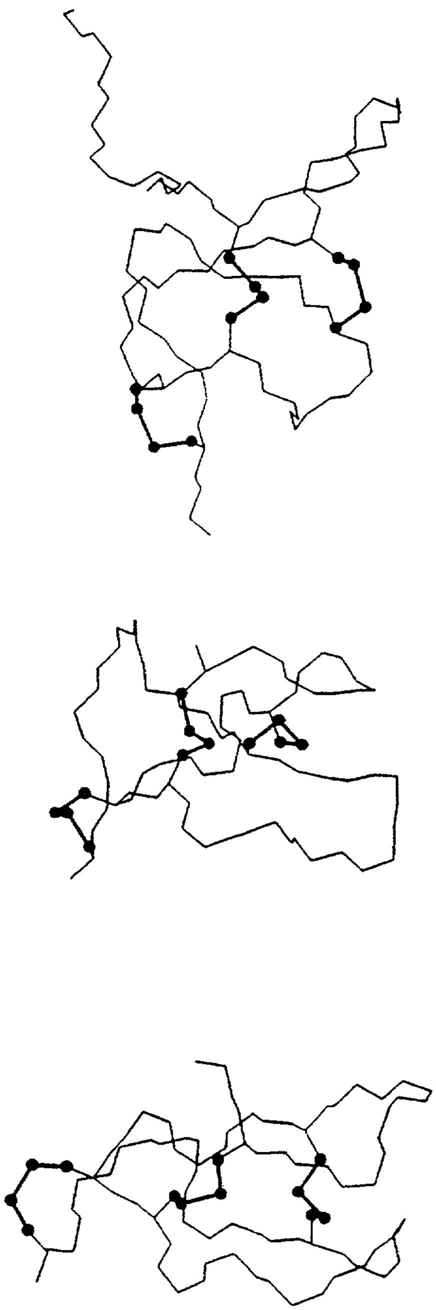
14. Use of a cyclic conotoxin peptide having activity at ion channel receptors as a neuropharmacological probe.

15. A method for the treatment or prophylaxis of conditions or diseases in mammals including the step of administering a cyclic conotoxin peptide.

16. Use of a cyclic conotoxin peptide in the manufacture of a medicament for the treatment or prophylaxis of diseases or conditions of mammals.

17. A composition comprising a cyclic conotoxin peptide and a pharmaceutically acceptable carrier or diluent.

18. A composition according to claim 17 which is a pharmaceutical composition.



GS

MVIIA

PVIIA

FIGURE 1

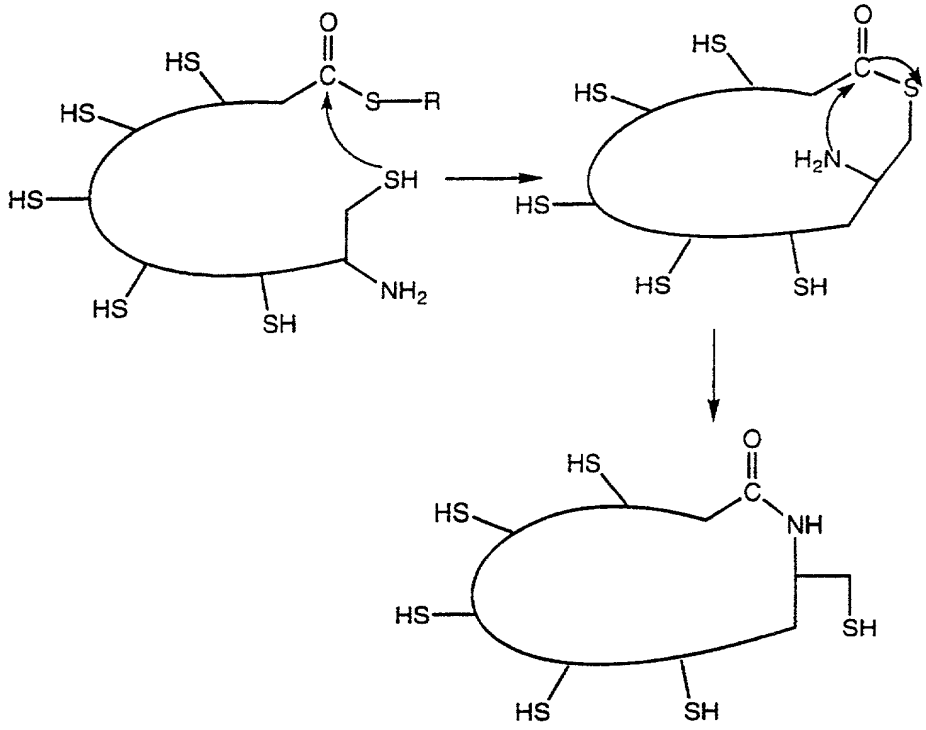


FIGURE 2

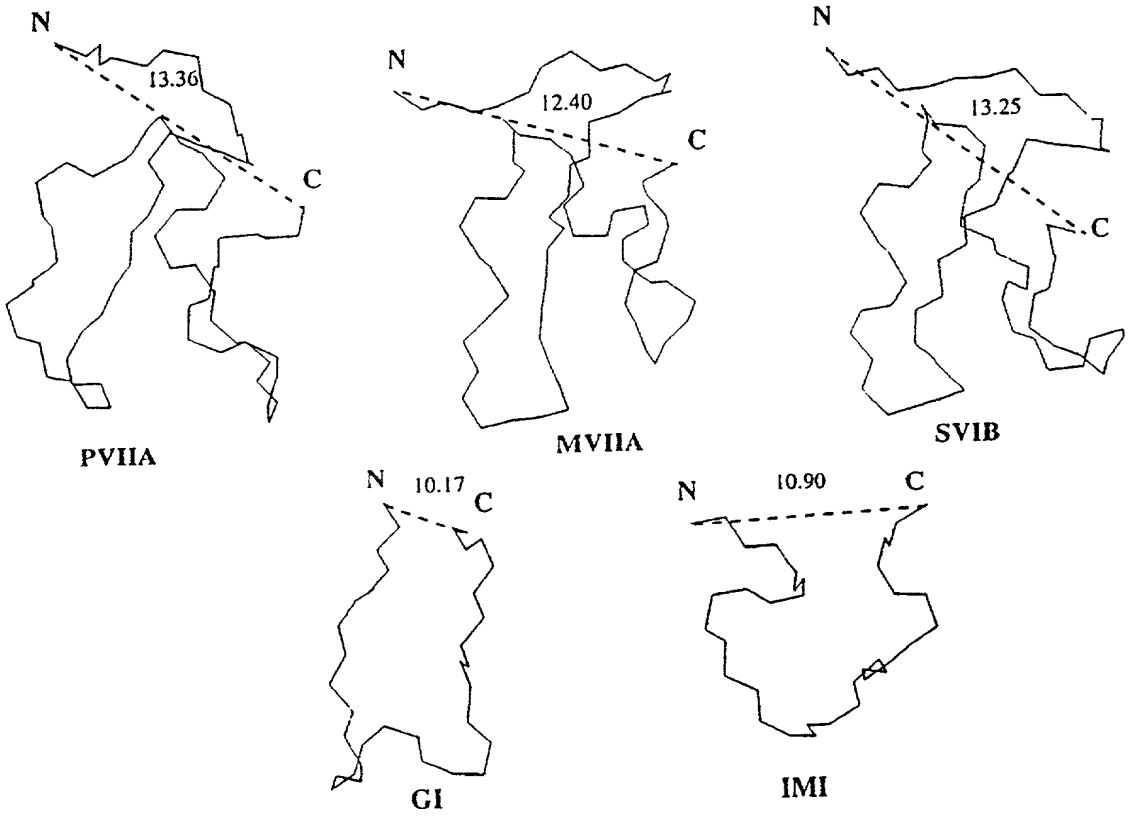


FIGURE 3

DOCKET NO. DAVI-0005 **PATENT**

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In Re U.S. National Phase Application of:
David James CRAIK; Norelle Lee DALY; and,
Katherine Justine Nielsen

Group Art Unit: Not yet known

International Serial No. PCT/AU99/00769

Examiner: Not yet assigned

International Filing Date: 14 September 1999

For: CYCLISED CONOTOXIN PEPTIDES

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name; and

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a

☒ Utility Patent ☐ Design Patent

is sought on the invention, whose title appears above, the specification of which:

☐ is attached hereto.

☒ was filed on 14 September 1999 as International Application No. PCT/AU99/00769 .

☒ said application having been amended on 8 June 2000 .

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to be material to the patentability of this application in accordance with 37 CFR § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a-d) of any **foreign application(s)** for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of any application on which priority is claimed:

Priority Claimed (If X'd)	**Country**	**Serial Number**	**Date Filed**
☒	AU	PP 5895	14 September 1998
☐			

I hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to be material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Serial Number	**Date Filed**	**Patented/Pending/Abandoned**

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below:

Serial Number	**Date Filed**

DOCKET NO. DAVI-0005 **PATENT**

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In Re Application of:

David James CRAIK, Norelle Lee DALY, and
Katherine Justine NIELSEN

Group Art Unit: Not yet known

International Application No.: PCT/AU99/00769

Examiner: Not yet assigned

International Filing Date: 14 September 1999

For: CYCLISED CONOTOXIN PEPTIDES

Assistant Commissioner for Patents
Washington DC 20231

Sir:

ASSOCIATE POWER OF ATTORNEY

The undersigned, of the firm WOODCOCK WASHBURN KURTZ MACKIEWICZ & NORRIS LLP, One Liberty Place - 46th Floor, Philadelphia, Pennsylvania 19103, Attorney and/or Agents for Applicant(s), hereby appoints the following:

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Michael P. Dunnam	Registration No. 32,611	Maureen S. Gibbons	Registration No. 44,121
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Albert J. Marcellino	Registration No. 34,664	Paul B. Milcetic	Registration No. 46,261
David R. Bailey	Registration No. 35,057	Joseph R. Condo	Registration No. 42,431
Doreen Yatko Trujillo	Registration No. 35,719	Michael K. Jones	Registration No. 41,100
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David A. Cherry	Registration No. 35,099	Eric H. Vance	Registration No. 47,151
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Norman L. Norris	Registration No. 24,196	Thomas E. Watson	Registration No. 43,243
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Date: 7/3/01

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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Mailing Address: 2/49 Bishop Street St. Lucia Queensland 4067 AUX Australia	**Date of Signature:** 21/5/01
City/State of Actual Residence: St. Lucia, Queensland 4067, Australia	**Citizenship:** Australia

Name: 3 of 3 KATHERINE JUSTINE NIELSEN	Katie Nielsen **Signature**
Mailing Address: 17 Lambourn Street Chapel Hill Queensland 4069 AUX Australia	**Date of Signature:** 11/5/01
City/State of Actual Residence: Chapel Hill, Queensland 4069, Australia	**Citizenship:** Australia

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